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Preclinical irradiations at conventional and very high dose rates (FLASH-RT) on small animals at the ARRONAX research cyclotron: repositioning, margins and field sizes



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Context

In the framework of translational research, it is envisioned to irradiate mice on radiobiology platforms, as the one of the ARRONAX cyclotron. In general, these facilities do not feature an integrated imaging system, which makes irradiation of organs without external anatomical landmarks challenging. Moreover, the field size cannot be tailored to each animal but must be set for a cohort.

This study aims to address the parameters required to determine the optimal field size for a selected cohort, which can be expressed as:

 $\label{eq:optimal Field size (cohort) = Field size (cohort) + \mbox{ Margins }_{\mbox{Targeting}} + \mbox{ Margins }_{\mbox{Respiratory}}$



1 Targets dimensions and inter-animal variability

The volumes of the boxes containing the **brain**, the **lungs** and the **intestines** were measured on 3x3 distinct tomographic images (XRAD225Cx, μ CBCT, 40kVp, 0.2mm isotropic voxel size) for **11 mice** (C57BL/6 - 5 females, 6 males) and compared to the animal's weight, as shown in Fig. 1. The relative uncertainty on the volume is the quadratic sum of the uncertainty of each length, corresponding to the size of two pixels.

- Brain volume was found to be independent of the weight
 Unique field size
- Lungs and intestinal volumes are correlated with the mouse weight
- → Selection of a cohort on the weight criterion enables a unique field size

2) Immobilization and targeting system

The system, shown in Fig. 2, consists of a patient-like **3D-printed bed** and harmless contentions, together with **graduations** as external markers for the lasers.

The targeting accuracy was assessed by resetting 10 times an anaesthetized C57BL/6 mouse for the **brain**, **lungs** and **intestines**. Each time, a μ CBCT (40kVp, 0.2mm isotropic voxel size) was acquired using the XRAD225Cx. The TPS μ Raystation® was used for **organ contouring** on the images and to perform rigid registrations, without rotation, with respect to the reference image.

The **maximum offsets** in the three directions were recorded in the **Table 1**. To ensure the target coverage, equivalent margins should be applied for each organ. An example of the histogram of deviations obtained for a site is also shown in Fig. 3.

	Right/Left	Longitudinal	Antero-Posterior
Brain	1, 4	0, 6	0, 9
Lungs	2, 0	3, 4	0, 4
Intestines	2, 3	3, 0	0, 7

TABLE 1 – Maximum offsets with respect to the reference image (mm)

3) Respiratory motion

In addition to the accuracy of the targeting, margins should also take into account potential **organ motions** such as respiratory motion. The **fluoroscopic** mode of the XRAD225Cx (40kVp, 0.1 mm voxel size) was used to acquire 10 respiratory cycles for a mouse. The wider cycle is displayed in Fig. 4.

Measured maximum amplitudes (full insipiration) relatives to full expiration position (Fig. 5.) are:

- 1.7 mm in the longitudinal direction (diaphragm)
- 0.6 mm in the antero-posterior direction.





Figure 4. Maximal amplitude cycle found with respect to time. Interpolation with Akima method.

Figure 5. Fluoroscopic mouse views.

Moreover, the respiration duration, which lasts 2.3 s in total, is long in comparison with FLASH-RT irradiation time (few hundred milliseconds) [1]. Then, the above amplitudes must be added to the margins to ensure the lungs coverage, where a loss could occur for 30 % of the irradiation time in conventional dose rate and up to 100 % of the time in FLASH-RT.

Conclusion and perspectives

- The margins obtained and the study of inter-animal variability guarantee optimal field sizes for irradiation of the brain, lungs and intestine.
- The methodology developed ensures accurate and reproducible positioning for targeted preclinical irradiations at ARRONAX.

Following steps will involve Monte Carlo GATE simulations to:

- Evaluate of the dosimetric impact of the respiratory cycle using the 4D digital mouse fantom (MOBY) [2],
- Verify the organ coverage, as well as distal and lateral dose homogeneity, on the cohort with adapted field size and margins found in this study.



(3)

Figure 1. For the three locations, volume containing the organ with respect to the animal weight.



Figure 2. Photograph of the mouse holding and targeting system.



Figure 3. Histogram of brain offsets measured on the 10 images.